

## Short and Convergent Synthesis of (1→3)-C-Linked Imino Disaccharides (Aza-C-disaccharides)

Yao-Hua Zhu and Pierre Vogel\*

Section de chimie, Université de Lausanne,  
BCH, CH-1015 Lausanne-Dorigny, Switzerland

Received September 1, 1998

Glycosidases and glycosyltransferases are key enzymes in the biosynthesis and processing of glycoproteins, which are molecules involved in recognition (cell–cell, host–pathogen interactions) and in control of biological mechanisms and structures.<sup>1</sup> Thus, substances able to inhibit these enzymes have become potential antibacterial, antiviral, antimetastatic, antidiabetes, antihyperglycemic, antiadhesive, or immunostimulatory agents.<sup>2,3</sup> A new class of selective glycosidase inhibitors has emerged with the C-linked imino disaccharides (aza-C-disaccharides)<sup>4,5</sup> which contains not only the steric and charge information of the glycosyl moiety, which is liberated during the enzyme-catalyzed hydrolysis, but also that of the aglycon to which the glycon is attached. The first example of a C-linked imino disaccharide (1,5-dideoxy-1,5-imino-D-mannitol linked at C(6) of D-galactose through a CH<sub>2</sub> unit) was prepared by Johnson and co-workers.<sup>6</sup> Other examples of “linear” C-linked imino disaccharides were obtained by the groups of Martin<sup>7</sup> and Van Boom.<sup>8</sup>

We have prepared the first examples of “branched” disaccharides.<sup>4,9</sup> Further examples were reported by Johnson and co-workers.<sup>5</sup> The current syntheses of “branched” disaccharide mimetics are rather lengthy, which limits their development and the search for better glycosidase inhibitors from among highly diverse candidates. We report here a new approach which condenses a dideoxyiminoaldose with an isolevoglucosone-derived enolate.<sup>10</sup> The aldol so obtained can be transformed readily into C-linked iminodisaccharides in which L-glycero-1,2,4-trideoxy-1,4-iminotetritol is linked through a CH(OH) moiety at position C(3) of methyl D-gluco- or methyl D-galacto-pyranoside.

Conjugate addition of benzyl alcohol to isolevoglucosone **1** (derived in four steps from D-glucose<sup>11</sup>) afforded ketone **2** in 90% yield with high stereoselectivity. The enolates of ketone **2** did not induce  $\beta$ -elimination at low temperature, probably because of the rigid bicyclic system.<sup>10</sup> The dichlorozinc enolate of **2**, generated by deprotonation with (Me<sub>3</sub>Si)<sub>2</sub>NK followed by treatment with anhydrous ZnCl<sub>2</sub>, added to aldehyde **3**.<sup>12</sup> This produced a single aldol **4** which partially decomposed during attempts at purification (chromatography on silica gel or Florisil).<sup>13,14</sup> Therefore, the crude aldol **4** was directly reduced with L-Selectride into the D-galactose derivative **5** (53% based on **2**).<sup>15</sup> Desilylation of **5** with *n*-Bu<sub>4</sub>NF afforded triol **6** in 96% yield. Debonylation of **6** gave **7** in 96% yield, which was refluxed in anhydrous methanol saturated with gaseous HCl, producing a 3:2 mixture of methyl  $\alpha$ - and  $\beta$ -D-galacto-pyranosides **8 $\alpha$**  and **8 $\beta$** , respectively, in 82% yield.

Reduction of the crude aldol **4** with Me<sub>4</sub>NBH(OAc)<sub>3</sub>,<sup>16</sup> on the other hand, formed the D-glucose derivative **9** (59% based on **2**) which was deprotected as above, giving first **10** in 94% yield, then **11** in 93% yield, and finally a 7:3 mixture of methyl  $\alpha$ - and  $\beta$ -D-gluco-pyranosides **12 $\alpha$**  and **12 $\beta$** , respectively, in 90% yield.

The structures of the new compounds were given by their mode of formation and their spectral data. Further proof was given by the vicinal coupling constants and the NOE's (Figure 1) measured in the <sup>1</sup>H NMR spectra of acetonides **13** and **14** derived from diols **5** and **9**, respectively, under standard conditions (Me<sub>2</sub>C(OMe)<sub>2</sub>, acetone, TsOH, and Drierite). The  $\delta_{C(Me)}$  = 24.8 and 24.2 ppm observed in the <sup>13</sup>C NMR spectrum of **13** suggest a twist-boat conformation for the acetonide.<sup>17</sup> Coupling

(1) See, e.g.: (a) Schauer, R. *Adv. Carbohydr. Chem. Biochem.* **1982**, *40*, 131. (b) Stütz, A. E. *Angew. Chem., Int. Ed. Engl.* **1996**, *35*, 1926. (c) Rademacher, T. W.; Parekh, R. B.; Dwek, R. A. *Annu. Rev. Biochem.* **1988**, *57*, 785. (d) Hindsgaul, O.; Khare, D. P.; Bach, M.; Lemieux, R. U. *Can. J. Chem.* **1985**, *63*, 2653. (e) Feizi, T. *Carbohydrate Recognition in Cellular Function*, Ciba Key Symposium 145; Wiley: Chichester, 1989; p 62. (f) Kornfeld, R.; Kornfeld, S. *Annu. Rev. Biochem.* **1985**, *54*, 631. (g) Moremen, K. W.; Trimble, R. B.; Herscovics, A. *Glycobiology* **1994**, *4*, 113. (h) Rice, G. E.; Bevilacqua, M. P. *Science* **1989**, *246*, 1303. (i) Smith, C. A.; Davis, T.; Anderson, D.; Solam, L.; Beckmann, M. P.; Jerzy, R.; Dower, S. K.; Cosman, D.; Goodwin, R. G. *Science* **1990**, *248*, 1019. (j) Varki, A. *Glycobiology* **1993**, *3*, 97. (k) Dwek, R. A. *Chem. Rev.* **1996**, *96*, 683.

(2) (a) Ganem, B. *Acc. Chem. Res.* **1996**, *29*, 340 and references cited therein. (b) van de Broek, L. A. G. M.; Vermaas, D. J.; Heskamp, B. M.; van Boeckel, C. A. A.; Tan, M. C. A. A.; Bolscher, J. G. M.; Ploegh, H. L.; van Kemenade, F. J.; de Goede, R. E. Y.; Miedema, F. *Recl. Trav. Chim. Pays-Bas* **1993**, *112*, 82. (c) Look, G. C.; Fotsch, C. H.; Wong, C.-H. *Acc. Chem. Res.* **1993**, *26*, 182. (d) Winchester, B.; Fleet, G. W. *J. Glycobiology* **1992**, *2*, 199. (e) Zheng, Y.; Pan, Y. T.; Asano, N.; Nash, R. J.; Elbein, A. D. *Glycobiology* **1997**, *7*, 297 and references cited therein.

(3) See, e.g.: (a) Pal, R.; Hoke, G. M.; Sarngadharan, M. G. *Proc. Natl. Acad. Sci. U.S.A.* **1989**, *86*, 3384. (b) Johnson, V. A.; Walker, B. D.; Barlow, M. A.; Paradis, T. J.; Chou, T. C.; Hirsch, M. S. *Antimicrob. Agents Chemother.* **1989**, *33*, 53. (c) Mehta, A.; Lu, X.; Block, T. M.; Blumberg, B. S.; Dwek, R. A. *Proc. Natl. Acad. Sci. U.S.A.* **1997**, *94*, 1822. (d) Fischer, P. B.; Karlsson, G. B.; Dwek, R. A.; Platt, F. M. *J. Virol.* **1996**, *70*, 7153. (e) Fenouillet, E.; Papandreou, M. J.; Jones, I. M. *Virology* **1997**, *231*, 89. (f) White, S. L.; Nagai, T.; Akijama, S. K.; Reeves, E. J.; Grzegorzewski, K.; Olden, K. *Cancer Commun.* **1991**, *3*, 83. (g) Olden, K.; Breton, P.; Grzegorzewski, K.; Yasuda, Y.; Gause, B. L.; Oredipe, O. A.; Newton, S. A.; White, S. L. *Pharmacol. Ther.* **1991**, *50*, 285. (h) Goss, P. E.; Baptiste, J.; Fernandes, B.; Baker, M.; Dennis, J. W. *Cancer Res.* **1994**, *54*, 1450.

(4) (a) Kraehenbuehl, K.; Picasso, S.; Vogel, P. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 893; (b) *Helv. Chim. Acta* **1998**, *81*, 1439.

(5) Johns, B. A.; Pan, Y. T.; Elbein, A. D.; Johnson, C. R. *J. Am. Chem. Soc.* **1997**, *119*, 4856.

(6) Johnson, C. R.; Miller, M. W.; Golebiowski, A.; Sundram, H.; Ksehati, M. B. *Tetrahedron Lett.* **1994**, *35*, 8991.

(7) Martin, O. R.; Liu, L.; Yang, F. *Tetrahedron Lett.* **1996**, *37*, 1991.

(8) Leeuwenburgh, M. A.; Picasso, S.; Overkleeft, H. S.; van der Marel, G. A.; Vogel, P.; Van Boom, J. H. Manuscript in preparation.

(9) (a) Baudat, A.; Vogel, P. *Tetrahedron Lett.* **1996**, *37*, 483. (b) Frérot, E.; Marquis, C.; Vogel, P. *Tetrahedron Lett.* **1996**, *37*, 2023. (c) Baudat, A.; Vogel, P. *J. Org. Chem.* **1997**, *62*, 6252.

(10) This approach has been reported by us for the synthesis of (1→3)-C-disaccharides: Zhu, Y.-H.; Vogel, P. *Tetrahedron Lett.* **1998**, *39*, 31.

(11) Horton, D.; Roski, J. P.; Norris, P. *J. Org. Chem.* **1996**, *61*, 3783.

(12) Mori, S.; Ohno, T.; Harada, H.; Aoyama, T.; Shioiri, T. *Tetrahedron* **1991**, *47*, 5051.

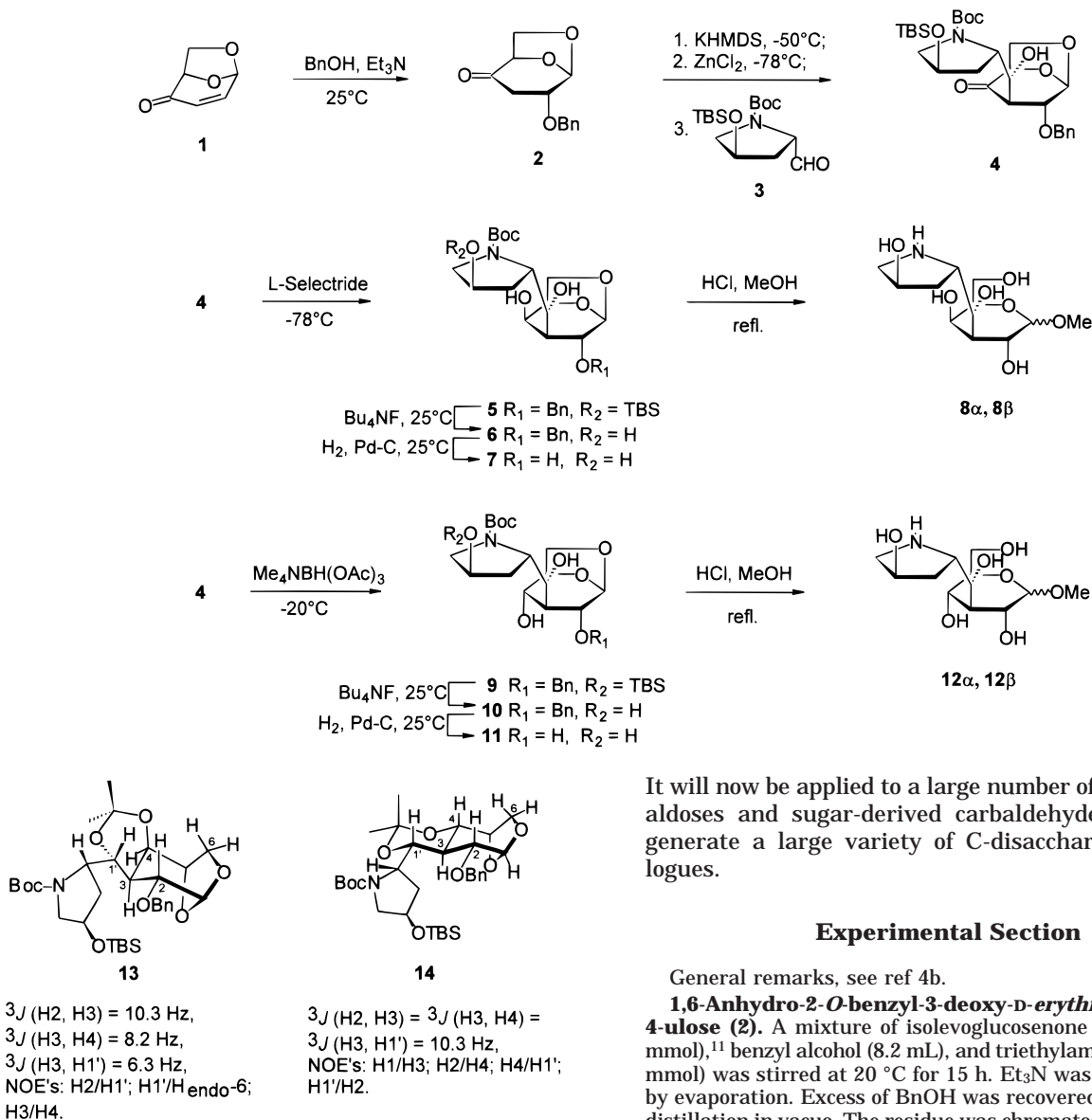
(13) Zinc enolates gave the best results among the enolates tried. Applying lithium enolates, retro-aldol products were formed during aqueous workup or during in situ reduction of the aldols (DIBALH, -78 °C); with aluminum enolates,  $\beta$ -elimination of BnOH took place during in situ reduction of the aldols (DIBALH, -78 °C).

(14) For examples of cross-aldolisations with sugar-derived ketones, see, e.g.: Yu, K. L.; Handa, S.; Tsan, R.; Fraser-Reid, B. *Tetrahedron* **1991**, *47*, 189.

(15) No formation of diol **9** was observed; NaBH<sub>4</sub> (THF/MeOH, 0 °C) gave a 10:1 mixture of diols **5** and **9**.

(16) Evans, D. A.; Chapman, K. T.; Carreira, E. M. *J. Am. Chem. Soc.* **1988**, *110*, 3560.

## Scheme 1



**Figure 1.** Selected  $^1\text{H}$  NMR data for **13** and **14**.

constants between vicinal protons at C(2,3,4) suggest a boat conformation for the tetrahydropyran ring (O–C1–C2–C3–C4–C5) in which H-2 and H-3 are axial. For **14**, the  $^{13}\text{C}$  NMR spectrum displays  $\delta_{\text{C}(\text{Me})} = 29.9$  and 19.9 ppm for the acetonide moiety, suggesting a chair conformation for it. Coupling constants between vicinal protons at C2, C3, and C4 also confirm a boat conformation for the tetrahydropyran ring as shown in Figure 1. This also appears to be the case for the 1,6-anhydro- $\beta$ -glucopyranose derivatives **9**, **10**, and **11** for which vicinal coupling constants  $^3J(\text{H}_3, \text{H}_4) = 8.7, 9.8,$  and  $10.5 \text{ Hz}$ , respectively, were measured. These data confirmed that the cross-alcoholization between **2** and **3** follows the Zimmerman–Traxler model.<sup>10,18</sup>

An expedient method for the stereoselective synthesis of (1→3)-C-linked imino disaccharides has been realized.

It will now be applied to a large number of iminodideoxy-aldoses and sugar-derived carbaldehydes in order to generate a large variety of C-disaccharides and analogues.

### Experimental Section

General remarks, see ref 4b.

**1,6-Anhydro-2-O-benzyl-3-deoxy-D-erythro-hexopyranose (2).** A mixture of isolevoglucosenone **1** (185 mg, 1.47 mmol),<sup>11</sup> benzyl alcohol (8.2 mL), and triethylamine (0.1 mL, 0.72 mmol) was stirred at 20 °C for 15 h. Et<sub>3</sub>N was then eliminated by evaporation. Excess of BnOH was recovered by bulb-to-bulb distillation in vacuo. The residue was chromatographed on silica gel (15:85 EtOAc/light petroleum ether), affording a colorless syrup (310 mg, 90%).  $[\alpha]_{\text{D}}^{25} = -57$  (c 1.2, CHCl<sub>3</sub>). IR (film): 1736 cm<sup>-1</sup>.  $^1\text{H}$  NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.45–7.22 (m, 5H), 5.62 (s, 1H), 4.63 and 4.57 (2d,  $J = 12.0 \text{ Hz}$ , 2H), 4.55 (d,  $J = 5.0 \text{ Hz}$ , 1H), 3.92 (d,  $J = 8.0 \text{ Hz}$ , 1H), 3.84 (dd,  $J = 8.0, 5.0 \text{ Hz}$ , 1H), 3.78 (ddd,  $J = 6.5, 3.5, 1.2 \text{ Hz}$ , 1H), 2.68 (dd,  $J = 12.5, 6.5 \text{ Hz}$ , 1H), 2.61 (dd,  $J = 12.5, 3.5 \text{ Hz}$ , 1H).  $^{13}\text{C}$  NMR (100.6 MHz, CDCl<sub>3</sub>):  $\delta$  204.1, 137.2, 128.6, 128.1, 127.7, 101.5, 79.1, 75.5, 71.6, 66.8, 39.2. MS (CI, NH<sub>3</sub>): 252 ([M + NH<sub>4</sub>]<sup>+</sup>). Anal. Calcd for C<sub>13</sub>H<sub>14</sub>O<sub>4</sub>: C, 66.66; H, 6.02. Found: C, 66.56; H, 6.01.

**1,6-Anhydro-2-O-benzyl-3-deoxy-3-[(1'R)-4'-O-tert-butylidimethylsilyl-N-tert-butyloxycarbonyl-2',3',5'-trideoxy-2',5'-imino-L-erythro-pentitol-1'-C-yl]- $\beta$ -D-galactopyranose (5).** A solution of potassium bis(trimethylsilyl)amide (0.75 M in toluene, 0.24 mL, 0.18 mmol) was added to a solution of **2** (32 mg, 0.14 mmol) in anhydrous THF (1.3 mL) stirred at –50 °C under a nitrogen atmosphere. After being stirred at this temperature for 1 h, the mixture was cooled to –78 °C and a 1 M solution of ZnCl<sub>2</sub> in ether (0.18 mL, 0.18 mmol) was added dropwise. After being stirred for 15 min, a solution of aldehyde **3**<sup>12</sup> (58 mg, 0.18 mmol) in anhydrous THF (0.3 mL) was added. The reaction mixture was warmed to –45 °C and stirred at this temperature for 5 h. A saturated aqueous solution of NH<sub>4</sub>Cl (0.6 mL) was added, and the mixture was allowed to warm to 20 °C with stirring. EtOAc (2 mL) was added. The aqueous phase was separated and extracted with EtOAc (3 mL, three times). The combined organic phases were dried (anhydrous Na<sub>2</sub>SO<sub>4</sub>), and

(17) (a) Buchanan, J. G.; Chacón-Fuertes, M. E.; Edgar, A. R.; Moorhouse, S. J.; Rawson, D. I.; Wightman, R. H. *Tetrahedron Lett.* **1980**, 21, 1793. (b) Rychnovsky, S. D.; Rogers, B.; Yang, G. *J. Org. Chem.* **1993**, 58, 3511.

(18) Zimmerman, H. E.; Traxler, M. D. *J. Am. Chem. Soc.* **1957**, 79, 1920.

the solvent was removed in vacuo to yield the crude aldol product **4**, the  $^1\text{H}$  NMR spectrum of which showed 75% conversion. This crude product was used for the following reaction without further purification.

A solution of L-Selectride (1 M in THF, 0.69 mL, 0.69 mmol) was added dropwise to a THF (2 mL) solution of above crude product at  $-78^\circ\text{C}$  under a nitrogen atmosphere. The resulting yellow solution was stirred at this temperature for 1 h. After being warmed to  $20^\circ\text{C}$ , MeOH (0.5 mL), followed by 3 M aqueous NaOH (1 mL) and 35%  $\text{H}_2\text{O}_2$  (1 mL), was added to the reaction mixture. The mixture was stirred vigorously at  $20^\circ\text{C}$  for 1 h. MeOH (2 mL) was added, and the mixture was extracted with  $\text{CH}_2\text{Cl}_2$  (6 mL, three times). The combined organic phases were dried (anhydrous  $\text{Na}_2\text{SO}_4$ ), and the solvent was removed in vacuo. The residue was purified by flash chromatography on silica gel (35:65 EtOAc/light petroleum ether), giving a colorless oil (41 mg, 53%).  $[\alpha]_D^{25} = -29$  (*c* 1.73,  $\text{CHCl}_3$ ). IR (film): 3364, 1673  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ , 323 K):  $\delta$  7.37–7.15 (m, 5H), 5.40 (s, 1H), 4.60 (s, 2H), 4.54 (dd, *J* = 5.8, 4.8 Hz, 1H), 4.51 (t, *J* = 6.1 Hz, 1H), 4.43–4.28 (m, 3H), 4.20 (m, 1H), 3.62–3.50 (m, 2H), 3.40 (d, *J* = 4.5 Hz, 1H), 3.23 (dd, *J* = 11.4, 4.2 Hz, 1H), 2.15 (ddd, *J* = 13.0, 7.9, 5.1 Hz, 1H), 2.05 (ddd, *J* = 7.6, 6.1, 4.5 Hz, 1H), 1.89 (dddd, *J* = 13.0, 7.7, 3.0, 1.6 Hz, 1H), 1.43 (s, 9H), 0.88 (s, 9H), 0.05 (s, 6H).  $^{13}\text{C}$  NMR (100.6 MHz,  $\text{CDCl}_3$ , 323 K):  $\delta$  156.3, 137.8, 128.6, 128.0, 127.7, 100.9, 80.5, 77.2, 74.7, 73.3, 71.6, 70.5, 66.6, 62.8, 60.7, 56.3, 41.0, 35.4, 28.4, 25.8, 18.0, -4.7, -4.8. MS (CI,  $\text{NH}_3$ ): 566 ( $[\text{M} + \text{H}]^+$ ). Anal. Calcd for  $\text{C}_{29}\text{H}_{47}\text{O}_8\text{NSi}$ : C, 61.56; H, 8.37; N, 2.48. Found: C, 61.66; H, 8.42; N, 2.45.

**1,6-Anhydro-2-O-benzyl-3-deoxy-3-[(1'R)-N-tert-butyl-oxycarbonyl-2',3',5'-trideoxy-2',5'-imino-L-erythro-pentitol-1'-C-yl]- $\beta$ -D-galactopyranose (6).** A tetrabutylammonium fluoride solution (1 M in THF, 0.40 mL, 0.40 mmol) was added to a solution of **5** (60 mg, 0.11 mmol) in THF (2.5 mL) at  $20^\circ\text{C}$ . After being stirred at  $20^\circ\text{C}$  for 15 h, the solvent was evaporated and the residue purified by chromatography on silica gel (1% MeOH in EtOAc), affording a colorless oil (48 mg, 96%).  $[\alpha]_D^{25} = -17$  (*c* 0.48,  $\text{CHCl}_3$ ). IR (film): 3374, 1668  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ , 323 K):  $\delta$  7.40–7.25 (m, 5H), 5.38 (s, 1H), 4.62 (d, *J* = 11.9 Hz, 1H), 4.53 (d, *J* = 11.9 Hz, 1H), 4.54 (dd, *J* = 6.4, 4.8 Hz, 1H), 4.50 (m, 1H), 4.45 (m, 1H), 4.36 (m, 1H), 4.28 (d, *J* = 7.6 Hz, 1H), 4.07 (m, 1H), 3.60 (d, *J* = 11.8 Hz, 1H), 3.54 (dd, *J* = 7.3, 4.8 Hz, 1H), 3.35 (d, *J* = 4.8 Hz, 1H), 3.30 (dd, *J* = 11.8, 4.2 Hz, 1H), 2.22 (ddd, *J* = 13.6, 7.6, 4.8 Hz, 1H), 2.00 (ddd, *J* = 8.8, 6.4, 4.8 Hz, 1H), 1.81 (dddd, *J* = 13.3, 7.8, 3.0, 2.1 Hz, 1H), 1.43 (s, 9H).  $^{13}\text{C}$  NMR (100.6 MHz,  $\text{CDCl}_3$ , 323 K):  $\delta$  156.0, 137.8, 128.6, 128.0, 100.7, 80.7, 77.9, 74.6, 72.8, 71.7, 70.0, 66.3, 62.7, 60.3, 56.1, 41.1, 34.6, 28.5. MS (CI,  $\text{NH}_3$ ): 452 ( $[\text{M} + \text{H}]^+$ ). Anal. Calcd for  $\text{C}_{23}\text{H}_{33}\text{O}_8\text{N}\cdot\text{H}_2\text{O}$ : C, 58.84; H, 7.51; N, 2.98. Found: C, 58.96; H, 7.42; N, 3.35.

**1,6-Anhydro-3-deoxy-3-[(1'R)-N-tert-butyl-oxycarbonyl-2',3',5'-trideoxy-2',5'-imino-L-erythro-pentitol-1'-C-yl]- $\beta$ -D-galactopyranose (7).** A degassed mixture of **6** (48 mg, 0.10 mmol), 10% Pd on charcoal (12 mg, 0.011 mmol), and MeOH (5 mL) was stirred under  $\text{H}_2$  at  $20^\circ\text{C}$  for 15 h. The catalyst was filtered off and solvent evaporated in vacuo. The residue was chromatographed (5% MeOH in EtOAc) to afford a white solid (37 mg, 96%). Mp  $209\text{--}210^\circ\text{C}$ .  $[\alpha]_D^{25} = -47$  (*c* 0.75, MeOH). IR (KBr): 3495, 3397, 3254, 1650  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{OD}$ , 323 K):  $\delta$  5.22 (d, *J* = 1.2 Hz, 1H), 4.64 (m, 1H), 4.56–4.38 (m, 3H), 4.30 (d, *J* = 7.6 Hz, 1H), 4.22 (dd, *J* = 8.2, 5.7 Hz, 1H), 3.60 (m, 2H), 3.56 (d, *J* = 11.4 Hz, 1H), 3.40 (dd, *J* = 11.4, 4.2 Hz, 1H), 2.35 (dt, *J* = 13.3, 5.7 Hz, 1H), 1.95 (dddd, *J* = 13.3, 8.2, 3.3, 1.8 Hz, 1H), 1.96–1.88 (m, 1H), 1.52 (s, 9H).  $^{13}\text{C}$  NMR (100.6 MHz,  $\text{CD}_3\text{OD}$ , 323 K):  $\delta$  156.7, 103.9, 81.5, 76.8, 73.8, 70.7, 68.3, 63.9, 60.9, 56.5, 44.7, 34.4, 28.9. MS (CI,  $\text{NH}_3$ ): 362 ( $[\text{M} + \text{H}]^+$ ). Anal. Calcd for  $\text{C}_{16}\text{H}_{27}\text{O}_8\text{N}$ : C, 53.18; H, 7.53; N, 3.88. Found: C, 53.23; H, 7.64; N, 3.83.

**Methyl 3-Deoxy-3-[(1'R)-2',3',5'-trideoxy-2',5'-imino-L-erythro-pentitol-1'-C-yl]- $\alpha$ (and  $\beta$ )-D-galactopyranoside (8 $\alpha$ , 8 $\beta$ ).** A mixture of **7** (10.0 mg, 0.0277 mmol) and anhydrous MeOH saturated with gaseous HCl (1 mL) was refluxed under  $\text{N}_2$  for 20 h. After cooling, the solution was poured onto a column (5 cm length) of Dowex 50WX8 (100–200 mesh). The column was washed sequentially with MeOH (20 mL),  $\text{H}_2\text{O}$  (5 mL), and 5%  $\text{NH}_3\cdot\text{H}_2\text{O}$  (40 mL). The fractions containing products **8 $\alpha$**  and **8 $\beta$**  were concentrated in vacuo and purified by flash chroma-

tography on silica gel (5% aqueous  $\text{NH}_3$ :*t*-PrOH = 1:2.5) to afford a colorless oil (7.4 mg, 82%).  $[\alpha]_D^{25} = +38$  (*c* 0.15,  $\text{H}_2\text{O}$ ). IR (film): 3375  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR (400 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  4.72 (d, *J* = 3.6 Hz, 0.6H, H-1 of **8 $\alpha$** ), 4.56 (m, 1H), 4.30 (d, *J* = 7.9 Hz, 0.4H, H-1 of **8 $\beta$** ), 4.24 (ddd, *J* = 10.3, 7.3, 4.5 Hz, 1H), 4.18 (m, 0.4 H), 4.15 (dd, *J* = 7.9, 4.5 Hz, 0.6 H), 4.12 (m, 0.6 H), 4.08 (d, *J* = 2.7 Hz, 0.4 Hz), 3.96 (dd, *J* = 11.5, 3.6 Hz, 0.6H), 3.82 (dd, *J* = 7.3, 4.8 Hz, 0.6H), 3.73–3.65 (m, 1.4H), 3.61 (dd, *J* = 11.2, 7.9 Hz, 0.4H), 3.55 (s, 1.2H), 3.41 (s, 1.8H), 3.18 (dd, *J* = 11.5, 4.5 Hz, 0.6 H), 3.15 (dd, *J* = 10.6, 4.5 Hz, 0.4H), 3.10 (d, *J* = 11.5 Hz, 0.6H), 3.07 (d, *J* = 10.6 Hz, 0.4H), 2.12 (dd, *J* = 13.3, 7.3 Hz, 1H), 2.04 (ddd, *J* = 13.3, 10.3, 4.2 Hz, 1H), 1.95 (ddd, *J* = 11.2, 7.9, 2.7 Hz, 0.6 H), 1.81 (ddd, *J* = 10.6, 6.0, 3.0 Hz, 0.4H).  $^{13}\text{C}$  NMR (100.6 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  106.5 (d,  $^1J(\text{C}, \text{H}) = 157$  Hz, C1 of **8 $\beta$** ), 99.9 (d,  $^1J(\text{C}, \text{H}) = 171$  Hz, C1 of **8 $\alpha$** ),<sup>19</sup> 79.0, 72.6, 71.8, 71.7, 71.3, 71.1, 68.0, 67.2, 67.1, 66.0, 62.4, 62.1, 61.7, 61.5, 58.3, 56.0, 54.3, 48.5, 44.0, 36.3, 35.9. MS (CI,  $\text{NH}_3$ ): 294 ( $[\text{M} + \text{H}]^+$ ). Anal. Calcd for  $\text{C}_{12}\text{H}_{23}\text{O}_7\text{N}\cdot 1.8\text{H}_2\text{O}$ : C, 44.25; H, 8.23; N, 4.30. Found: C, 44.26; H, 8.53; N, 4.44.

**1,6-Anhydro-2-O-benzyl-3-deoxy-3-[(1'R)-4'-O-tert-butyl-dimethylsilyl-N-tert-butyl-oxycarbonyl-2',3',5'-trideoxy-2',5'-imino-L-erythro-pentitol-1'-C-yl]- $\beta$ -D-glucopyranose (9).** A solution of crude aldol product **4** derived from 32 mg (0.14 mmol) of **2** (see above) in MeCN (0.3 mL) was added to a stirred solution of Me<sub>2</sub>NBH(OAc)<sub>3</sub> (330 mg, 1.3 mmol) in  $\text{CH}_3\text{CN}$  (1 mL) and AcOH (1 mL) at  $-40^\circ\text{C}$ . The flask containing **4** was rinsed with  $\text{CH}_3\text{CN}$  (0.3 mL, twice). The mixture was warmed to  $-20^\circ\text{C}$  and stirred at this temperature for 20 h. A 1 M aqueous potassium sodium tartrate solution (1 mL) was added, and the mixture was warmed to  $20^\circ\text{C}$ . The phases were separated. The aqueous phase was extracted with  $\text{CH}_2\text{Cl}_2$  (3 mL, three times). The combined organic phases were washed with a saturated aqueous solution of  $\text{NaHCO}_3$  (5 mL), which was counter-extracted with  $\text{CH}_2\text{Cl}_2$  (3 mL, four times). The washing and extraction procedure was repeated once more. The combined organic phases were dried (anhydrous  $\text{Na}_2\text{SO}_4$ ), and the solvent was evaporated in vacuo. Chromatography on silica gel (2:3 EtOAc/light petroleum ether) afforded a colorless oil (32 mg, 59%).  $[\alpha]_D^{25} = -32$  (*c* 1.5,  $\text{CHCl}_3$ ). IR (film): 3428, 1674  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ , 323 K):  $\delta$  7.39–7.26 (m, 5H), 5.42 (s, 1H), 4.60 (s, 2H), 4.49 (d, *J* = 5.2 Hz, 1H), 4.33 (p, *J* = 3.9 Hz, 1H), 4.14 (td, *J* = 7.5, 3.0 Hz, 1H), 3.98 (dd, *J* = 8.1, 2.6 Hz, 1H), 3.84 (m, 2H), 3.66 (dd, *J* = 7.3, 5.2 Hz, 1H), 3.55 (d, *J* = 11.5 Hz, 1H), 3.26 (dd, *J* = 11.5, 3.9 Hz, 1H), 3.16 (d, *J* = 3.9 Hz, 1H), 2.11 (ddd, *J* = 13.3, 7.7, 3.9 Hz, 1H), 1.93 (dddd, *J* = 13.3, 7.7, 3.9, 1.8 Hz, 1H), 1.87 (ddd, *J* = 8.7, 8.5, 3.9 Hz, 1H), 1.44 (s, 9H), 0.86 (s, 9H), 0.05 (s, 3H), 0.03 (s, 3H).  $^{13}\text{C}$  NMR (100.6 MHz,  $\text{CDCl}_3$ , 323 K):  $\delta$  156.1, 137.7, 128.5, 127.9, 127.7, 100.4, 80.3, 77.6, 77.3, 73.8, 71.3, 70.6, 68.9, 66.3, 59.7, 56.3, 45.4, 35.2, 28.5, 25.8, 18.0, -4.8, -4.9. MS (CI,  $\text{NH}_3$ ): 566 ( $[\text{M} + \text{H}]^+$ ). Anal. Calcd for  $\text{C}_{29}\text{H}_{47}\text{O}_8\text{NSi}$ : C, 61.56; H, 8.37; N, 2.48. Found: C, 61.51; H, 8.35; N, 2.44.

**1,6-Anhydro-2-O-benzyl-3-deoxy-3-[(1'R)-N-tert-butyl-oxycarbonyl-2',3',5'-trideoxy-2',5'-imino-L-erythro-pentitol-1'-C-yl]- $\beta$ -D-glucopyranose (10).** The same procedure as for the preparation of **6** was used, starting from **9** (119 mg, 0.211 mmol), yielding a colorless oil (93 mg, 94%) which crystallized from  $\text{CH}_2\text{Cl}_2$ . Mp  $80\text{--}81^\circ\text{C}$ .  $[\alpha]_D^{25} = -49$  (*c* 1.75, MeOH). IR (KBr): 3414, 1656  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{OD}$ , 323 K):  $\delta$  7.45–7.28 (m, 5H), 5.45 (s, 1H), 4.64 (s, 2H), 4.45 (m, 1H), 4.43 (m, 1H), 4.10–3.98 (m, 1H), 4.03 (d, *J* = 9.8 Hz, 1H), 3.93 (m, 1H), 3.82 (d, *J* = 7.6 Hz, 1H), 3.67 (dd, *J* = 7.6, 5.9 Hz, 1H), 3.52 (d, *J* = 11.3 Hz, 1H), 3.43 (dd, *J* = 11.3, 4.9 Hz, 1H), 3.22 (br s, 1H), 2.33 (br s, 1H), 1.89 (ddd, *J* = 13.3, 7.9, 3.0 Hz, 1H), 1.75 (dt, *J* = 9.8, 3.0 Hz, 1H), 1.44 (s, 9H).  $^{13}\text{C}$  NMR (100.6 MHz,  $\text{CD}_3\text{OD}$ , 323 K):  $\delta$  156.6, 139.3, 129.6, 129.1, 129.0, 101.5, 81.3, 78.8, 75.8, 73.1, 72.2, 70.8, 68.3, 66.3, 60.7, 56.2, 47.2, 34.4, 28.8. Anal. Calcd for  $\text{C}_{23}\text{H}_{33}\text{O}_8\text{N}\cdot\text{H}_2\text{O}$ : C, 58.84; H, 7.51; N, 2.98. Found: C, 59.16; H, 7.40; N, 2.73.

**1,6-Anhydro-3-deoxy-3-[(1'R)-N-tert-butyl-oxycarbonyl-2',5'-imino-2',3',5'-trideoxy-L-erythro-pentitol-1'-C-yl]- $\beta$ -D-glucopyranose (11).** The same procedure as for the preparation

(19) (a) Bock, K.; Thøgersen, H. *Annu. Rep. NMR Spectrosc.* **1982**, *13*, 1. (b) Bock, K.; Pedersen, C. *Adv. Carbohydr. Chem. Biochem.* **1983**, *41*, 27. (c) Jones, C. In *Advances in Carbohydrate Analysis*; JAI Press: Greenwich, CT, 1991; Vol. 1, p 145.



of **7** was used, starting from **10** (50 mg, 0.11 mmol): white solid (37 mg, 93%). Mp 164–165 °C.  $[\alpha]_D^{25} = -74$  (c 1.2, MeOH). IR (KBr): 3463, 3379, 1649  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{OD}$ , 323 K):  $\delta$  5.27 (s, 1H), 4.45 (d,  $J = 5.4$  Hz, 1H), 4.42 (m, 1H), 4.24 (m, 1H), 4.07 (dd,  $J = 10.4$ , 1.5 Hz, 1H), 3.92 (m, 1H), 3.82 (d,  $J = 7.6$  Hz, 1H), 3.67 (dd,  $J = 7.6$ , 5.4 Hz, 1H), 3.55 (d,  $J = 11.5$  Hz, 1H), 3.42 (dd,  $J = 11.5$ , 4.6 Hz, 1H), 3.40 (d,  $J = 3.1$  Hz, 1H), 2.32 (dt,  $J = 13.4$ , 6.0 Hz, 1H), 1.96 (dddd,  $J = 13.3$ , 8.1, 3.1, 1.5 Hz, 1H), 1.59 (dt,  $J = 10.5$ , 3.1 Hz, 1H), 1.52 (s, 9H).  $^{13}\text{C}$  NMR (100.6 MHz,  $\text{CD}_3\text{OD}$ , 323 K):  $\delta$  156.8, 104.2, 81.2, 78.8, 73.3, 70.8, 69.2, 68.8, 66.5, 60.6, 56.4, 49.8, 34.3, 28.9. MS (CI,  $\text{NH}_3$ ): 362 ( $[\text{M} + \text{H}]^+$ ). Anal. Calcd for  $\text{C}_{16}\text{H}_{27}\text{O}_8\text{N}$ : C, 53.18; H, 7.53; N, 3.88. Found: C, 53.02; H, 7.68; N, 3.83.

**Methyl 3-Deoxy-3-[(1'R)-2',5'-imino-2',3',5'-trideoxy-L-erythro-pentitol-1'-C-yl]- $\alpha$  (and  $\beta$ )-D-glucopyranoside (12 $\alpha$ , 12 $\beta$ )**. The same procedure as for the preparation of **8 $\alpha$**  and **8 $\beta$**  was used, starting from **11** (10.0 mg, 0.0277 mmol), chromatography on silica gel (5% aqueous  $\text{NH}_3$ :MeOH = 1:12): colorless oil (7.8 mg, 90%).  $[\alpha]_D^{25} = +43$  (c 0.29,  $\text{H}_2\text{O}$ ). IR (film): 3354  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR (400 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  4.71 (d,  $J = 3.7$  Hz, 0.7H, H-1 of **12 $\alpha$** ), 4.59 (t,  $J = 4.2$  Hz, 1H), 4.35 (d,  $J = 7.9$  Hz, 0.3H, H-1 of **12 $\beta$** ), 4.26 (dd,  $J = 6.0$ , 2.6 Hz, 1H), 4.14 (dt,  $J = 11.4$ , 6.0 Hz, 1H), 3.88 (dd,  $J = 12.1$ , 2.1 Hz, 0.3H), 3.83 (dd,  $J = 12.1$ , 2.1 Hz, 0.7H), 3.70 (d,  $J = 12.1$ , 0.3H), 3.67 (d,  $J = 12.1$ , 0.7H), 3.68–3.58 (m, 2H), 3.65 (dd,  $J = 10.6$ , 3.7 Hz, 0.7 H), 3.54 (s, 0.9H), 3.41 (s, 2.1H), 3.34 (dd,  $J = 12.8$ , 4.2 Hz, 1H), 3.28 (d,  $J = 7.7$ , 0.3H), 3.21 (d,  $J = 12.6$ , 1H), 2.18 (dd,  $J = 14.0$ , 6.0 Hz, 1H), 1.98 (ddd,  $J = 14.0$ , 11.4, 4.2 Hz, 1H), 1.96 (td,  $J = 10.9$ , 3.5 Hz, 0.7H), 1.80 (td,  $J = 10.3$ , 2.3 Hz, 0.3H).  $^{13}\text{C}$  NMR (100.6 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  105.7 (d,  $^1J(\text{C}, \text{H}) = 163$  Hz, C1 of **12 $\beta$** ), 99.4 (d,  $^1J(\text{C}, \text{H}) = 167$  Hz, C1 of **12 $\alpha$** ),<sup>19</sup> 79.3, 72.8, 70.8, 70.0, 69.9, 69.9, 68.1, 64.9, 64.5, 62.4, 62.1, 61.9, 61.7, 58.1, 55.9, 54.1, 49.5, 45.4, 37.2, 37.0. MS (CI,  $\text{NH}_3$ ): 294 ( $[\text{M} + \text{H}]^+$ ). Anal. Calcd for  $\text{C}_{12}\text{H}_{23}\text{O}_7\text{N} \cdot 1.2\text{H}_2\text{O}$ : C, 45.76; H, 8.13; N, 4.45. Found: C, 45.83; H, 7.99; N, 4.53.

**1,6-Anhydro-2-O-benzyl-3-deoxy-1',4-O-isopropylidene-3-[(1'R)-4'-O-tert-butylidimethylsilyl-N-tert-butyloxycarbonyl-2',3',5'-trideoxy-2',5'-imino-L-erythro-pentitol-1'-C-yl]- $\beta$ -D-galactopyranose (13)**. A mixture of **5** (39 mg, 0.069 mmol), 2,2-dimethoxypropane (2 mL), Drierite (2 g, powder), dry acetone (4 mL), and *p*-toluenesulfonic acid (1 mg) was stirred at 20 °C for 15 h. After addition of  $\text{K}_2\text{CO}_3$  (10 mg), the precipitate was filtered off and the solvent evaporated in vacuo. The residue was purified by chromatography on silica gel (8:92 EtOAc/light petroleum ether), affording a colorless oil (37 mg, yield 89%).  $[\alpha]_D^{25} = +5$  (c 1.65,  $\text{CHCl}_3$ ). IR (film): 1694  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR (400

MHz,  $\text{CDCl}_3$ , 333 K):  $\delta$  7.40–7.22 (m, 5H), 5.42 (s, 1H), 4.63 (s, 2H), 4.57 (dd,  $J = 7.9$ , 5.6 Hz, 1H), 4.45 (p,  $J = 5.2$  Hz, 1H), 4.38 (t,  $J = 8.2$  Hz, 1H), 4.36–3.98 (m, 2H), 4.22 (d,  $J = 7.5$ , 1H), 3.52 (dd,  $J = 7.5$ , 5.6 Hz, 1H), 3.41 (dd,  $J = 10.7$ , 5.4 Hz, 1H), 3.43–3.21 (m, 2H), 2.23 (dt,  $J = 12.6$ , 5.4 Hz, 1H), 1.90 (ddd,  $J = 10.3$ , 8.2, 6.3 Hz, 1H), 1.79 (ddd,  $J = 12.4$ , 8.6, 5.6 Hz, 1H), 1.49 (s, 9H), 1.32 (s, 3H), 1.31 (s, 3H), 0.88 (s, 9H), 0.06 (s, 3H), 0.04 (s, 3H).  $^{13}\text{C}$  NMR (100.6 MHz,  $\text{CDCl}_3$ , 333K):  $\delta$  154.3, 137.8, 128.4, 127.8, 127.7, 101.8, 80.4, 79.4, 71.3, 70.6, 70.5, 70.2, 62.5, 61.9, 57.3, 55.1, 40.0, 34.9, 28.7, 25.8, 24.8, 24.2, 18.0, –4.8, –4.9. MS (CI,  $\text{NH}_3$ ): 606 ( $[\text{M} + \text{H}]^+$ ). Anal. Calcd for  $\text{C}_{32}\text{H}_{51}\text{O}_8\text{NSi}$ : C, 63.44; H, 8.48; N, 2.31. Found: C, 63.53; H, 8.56; N, 2.41.

**1,6-Anhydro-2-O-benzyl-3-deoxy-1',4-O-isopropylidene-3-[(1'R)-4'-O-tert-butylidimethylsilyl-N-tert-butyloxycarbonyl-2',3',5'-trideoxy-2',5'-imino-L-erythro-pentitol-1'-C-yl]- $\beta$ -D-glucopyranose (14)**. The same procedure as for the preparation of **13** was used, starting from **9** (32 mg, 0.057 mmol): colorless oil (29 mg, 85%).  $[\alpha]_D^{25} = -11$  (c 1.45,  $\text{CHCl}_3$ ). IR (film): 1690  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ , 323 K):  $\delta$  7.40–7.28 (m, 5H), 5.32 (s, 1H), 4.60 (s, 2H), 4.44 (p,  $J = 5.6$  Hz, 1H), 4.42 (m, 1H), 4.23 (m, 1H), 4.18 (m, 1H), 3.63 (m, 1H), 3.48 (m, 2H), 3.38 (dd,  $J = 10.7$ , 5.4 Hz, 1H), 3.25 (m, 1H), 3.19 (m, 1H), 2.23 (ddd,  $J = 12.8$ , 5.6, 4.4 Hz, 1H), 1.77 (m, 1H), 1.60 (q,  $J = 10.3$  Hz, 1H), 1.46 (s, 9H), 1.39 (s, 6H), 0.86 (s, 9H), 0.03 (s, 3H), 0.01 (s, 3H).  $^{13}\text{C}$  NMR (100.6 MHz,  $\text{CDCl}_3$ , 323 K):  $\delta$  154.3, 137.5, 128.5, 127.9, 102.0, 79.0, 78.8, 75.7, 73.1, 71.7, 70.7, 69.3, 58.1, 55.1, 38.7, 34.5, 29.9, 28.6, 25.9, 19.9, 18.0, –4.8, –4.9. MS (CI,  $\text{NH}_3$ ): 606 ( $[\text{M} + \text{H}]^+$ ). Anal. Calcd for  $\text{C}_{32}\text{H}_{51}\text{O}_8\text{NSi}$ : C, 63.44; H, 8.48; N, 2.31. Found: C, 63.48; H, 8.46; N, 2.37.

**Acknowledgment.** We thank the Swiss National Science Foundation and the Fonds Herbette (Lausanne) for financial support. We thank also Mr. F. Sepulveda and Mr. Rey for their technical assistance.

**Supporting Information Available:** Detailed  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra and signal assignments, further optical rotation data, and UV, IR, and MS spectra (12 pages). This material is contained in libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.

JO981781D